

1 **A niche for isotopic ecology**

2

3 Seth D. Newsome¹, Carlos Martinez del Rio², Stuart Bearhop³, Donald L. Phillips⁴

4

5 ¹Carnegie Institution of Washington, Geophysical Laboratory, 5251 Broad Branch Road NW,
6 Washington, DC 20015, USA

7

8 ²Department of Zoology and Physiology, University of Wyoming, Laramie, WY 82071-3166,
9 USA

10

11 ³Centre for Ecology & Conservation, School of Biosciences, University of Exeter, Cornwall
12 Campus, Penryn, Cornwall TR10 9EZ, UK

13

14 ⁴U.S. Environmental Protection Agency. National Health & Environmental Effects Research
15 Laboratory. 200 SW 35th St. Corvallis, OR 97333, USA

16

17

18

19 **Running title**

20 Isotopic niche

21

22 **Keywords**

23 niche, stable isotopes, $\delta^{15}\text{N}$, $\delta^{13}\text{C}$

24

25 Abstract (152 words), 53 references, 1 table, and 5 figures

26

27 *12/4/06 Version for Submission, Frontiers in Ecology and the Environment*

28

29 **Abstract**

30 **Fifty years ago Hutchinson defined the ecological niche as a hypervolume in n-dimensional**
31 **space with environmental variables as axes. Ecologists have recently developed renewed**
32 **interest in the concept. Hutchinson divided factors defining the niche into two categories:**
33 **bionomic and scenopoetic. Technological advances now allow ecologists to use stable**
34 **isotope analyses to quantify these niche dimensions. Analogously, we define the isotopic**
35 **niche as an area (in δ -space) with isotopic values (δ -values) as coordinates. We illustrate the**
36 **isotopic niche with two examples: the ontogenetic niche and the application of historic**
37 **ecology to conservation biology. To make isotopic measurements comparable to other niche**
38 **formulations we propose transforming δ -space to p-space, where axes represent relative**
39 **proportions of isotopically distinct resources incorporated into an animal's tissues.**
40 **Sustaining renewed interest in the niche requires novel methods to measure variables that**
41 **define it. Stable isotope analyses are a natural, perhaps crucial, tool in contemporary**
42 **studies of the ecological niche.**

43
44 **In A Nutshell**

- 45 • Stable isotope analysis (SIA) provides quantitative information on both bionomic and
46 scenopoetic factors (axes) commonly used to define ecological niche space.
- 47 • Advances in isotope mixing models allow transforming isotopic data into source
48 contribution values, thus providing a standardized means to characterize an organism's
49 ecological niche.
- 50 • Implicit in this approach is a thorough understanding of the isotopic variation within and
51 among source pools available to consumers and the recognition that isotopic analysis
52 does not typically provide information on taxon-specific resource use.
- 53 • Careful implementation of SIA will benefit studies of resource competition in community
54 structure, as well as help characterize population-level biogeography or connectivity
55 crucial for successful conservation of highly migratory and/or elusive species.

56
57
58 **Introduction**

59 The term ecological niche is as fundamental to ecology as it is elusive. Niches are central to
60 ecological thinking because they represent convenient shorthand for many of the concepts that
61 ecologists use to approach a variety of important problems, which include resource use,
62 geographical diversity, and many aspects of community composition and structure (McGill et al.
63 2006). Niches are elusive for two reasons. First, there is not one, but many niche concepts, each
64 of which emphasizes a different aspect of a species' ecological characteristics (Leibold 1995).
65 The second reason for the elusiveness of the ecological niche is that it is difficult to measure. The

66 confusion and ambiguity that often surrounds the niche has led some ecologists to call for
67 purging the ecological literature of niches (Hubbell 2001). Indeed, until relatively recently, the
68 niche fell in disuse, and alternative terms replaced some of its traditional meanings (Chase and
69 Liebold 2003).

70 Yet the niche persists and seems to be making a striking comeback. As an example, the
71 niche was featured prominently in all the articles of a recent supplement of *Ecology* devoted to
72 phylogenetic approaches to community ecology (*Ecology*. 2006. 87(7)). Over the last few years,
73 niche definitions abandoned as inoperative have been renovated into relatively well-defined and
74 functional concepts. Joseph Grinnell's (1917) "habitat" concept of the niche has reincarnated
75 into the bioclimatic niche measured by geographical distribution area modelers (Elith et al.
76 2006). In a similar fashion, Elton's niche concept of the role of a species in a community has
77 morphed into Chase and Leibold's (2003) definition of the functional (or net-growth isocline,
78 NGI) niche. Both the bioclimatic niche and the functional/NGI niche owe their existence to
79 progress in analytical and computational methods, as well as to conceptual advances in ecology.
80 The bioclimatic niche relies heavily on the development of effective geographical information
81 technologies and on the ability of machines to handle large amounts of spatially explicit data
82 analyzed by computationally intensive models (Elith et al. 2006). The functional niche is
83 pivotally dependent on Tilman's (1988) concept of zero net growth isoclines (or ZNGIs, see
84 Chase and Leibold 2003). The niche concept that we develop here is similarly dependent on both
85 technological and conceptual advances.

86 We postulate the "isotopic niche" as a construct that can inform questions traditionally
87 considered within the broad domain of the ecological niche – including the functional and
88 bioclimatic niche concepts. We suggest that stable isotopes analyses (SIA) offer a superb tool to

89 assess many of the ecological characteristics of organisms that niche research aims to investigate.
90 In following sections we define the isotopic niche, and explain the kind of information that it can
91 disclose. Perhaps more importantly, we also identify the limitations of isotopic niches. Then, we
92 propose that the variation in isotopic incorporation among an animal's tissues permits
93 characterizing the contribution of intra- and inter-individual variation to a species isotopic niche.
94 We exemplify the utility of isotopic niches with two examples: the use of SIA to track changes in
95 the ecological characteristics of organisms through ontogeny, and as tools in conservation
96 biology. Finally, we describe the relationship between the isotopic niche and other niche
97 constructs and outline the transformations of the isotopic niche space that one must perform to
98 make the metrics of the isotopic niche comparable to those estimated in other formulations of the
99 ecological niche. Our discussion emphasizes animals, but our approach can be modified to
100 define botanical and microbiological isotopic niches as well.

101

102 **Delta spaces and the isotopic niche**

103 Almost 50 years ago, George Evelyn Hutchinson (1957) formalized the ecological niche as an
104 abstract n-dimensional set of points in a space whose axes represent environmental variables. In
105 subsequent elaborations of the niche, Hutchinson (1978) established a useful distinction between
106 scenopoetic and bionomic niche axes. The scenopoetic axes are those that set the bioclimatic
107 stage in which a species performs (Hutchinson 1978), whereas the bionomic axes are those that
108 define the resources that animals use. After Hutchinson's original formulation, the niche has
109 undergone many changes, but all alternative contemporary definitions retain the formalization of
110 the niche as a multidimensional space. Isotopic ecologists have been representing the results of

111 their analyses in niche-like multivariate spaces with coordinates that are analogous to
112 Hutchinson's scenopoetic and bionomic axes.

113 The analysis of stable isotopes has emerged as a key tool for ecologists (Fig. 1 and Table
114 1). Stable isotopes are useful because many physicochemical (i.e., kinetic reactions) and
115 biochemical processes (i.e., equilibrium reactions) are sensitive to differences in the dissociation
116 energies of molecules, which often depend on the mass of the elements from which these
117 molecules are made. Thus, the isotopic composition of many materials, including the tissues of
118 organisms, often contains a label of the process that created it. For example, the producers at the
119 base of food webs often imprint the biological molecules that they manufacture with distinct
120 carbon, nitrogen, and hydrogen signatures (Farquhar 1989, Robinson 2001). Because consumers
121 incorporate these "signatures" into their tissues, we can use $^{13}\text{C}/^{12}\text{C}$ and $^2\text{H}/^1\text{H}$ to identify their
122 reliance on producers with different photosynthetic pathways –i.e. C_3 , C_4 , or CAM (Wolf and
123 Martinez del Rio 2003). We can also use a combination of $^{13}\text{C}/^{12}\text{C}$ and $^{15}\text{N}/^{14}\text{N}$ to determine the
124 contribution of marine and terrestrial food webs to an animal's diet or estimate trophic position
125 (Post 2002 and references there). These are examples of the application of stable isotope
126 analyses to the elucidation of variables along bionomic axes. Stable isotopes can also give us
127 insight into the scenopoetic dimensions of the niche, such as environmental temperature or
128 habitat latitude (Table 1).

129 The term "isotopic fractionation" refers to the difference in isotopic composition between
130 the reactants and products of a physicochemical process. Isotopic fractionations can be
131 temperature dependent (Fry 2006), so the temperature at which a fractionating process takes
132 place is often recorded in the isotopic composition of the products. For example, the
133 temperature-dependent fractionation of oxygen during the synthesis of calcium carbonate

134 provides a convenient isotopic thermometer that measures the temperature at which permanent
135 carbonate-containing structures such as shells, otoliths, and bones are synthesized (Radtke et al.
136 1996). The isotopic composition of rainwater is determined by a combination of factors, which
137 include altitude, latitude, distance from the coast, and temperature. These factors create the
138 broadly predictable geographical patterns in the $\delta^{18}\text{O}$ and δD of precipitation (Bowen 2003).
139 These “isoscares” have been used widely to track animal movements (Rubenstein & Hobson
140 2004, Fig. 2). West et al. (2006) have aptly referred to stable isotopes as nature’s recorders of
141 ecological processes. Stable isotopes represent “wireless sensors” (sensu Collins et al. 2006) for
142 a variety of the bionomic and scenopoetic ecological variables that Hutchinson envisioned as
143 elements of the niche.

144 Isotopic ecologists often present their measurements as points in Cartesian spaces in
145 which axes represent the delta (δ) values for different elements (Fig. 1 and Fig. 3). This “ δ -
146 space” is closely related to the n-dimensional space that contains what ecologists refer to as the
147 niche. Indeed, isotopic ecologists have used delta spaces to explore questions that have been
148 traditionally within the domain of niche theory. For example, Genner et al. (1999) and Bocher et
149 al. (2000) used $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ values to document niche segregation in cichlids and petrels,
150 respectively.

151 SIA is particularly well suited to investigate the intra- and inter-individual components of
152 niche breadth. Because different animal tissues incorporate the isotopic signatures of resources at
153 different rates, they can integrate information over different temporal periods, which is a major
154 advantage of SIA in comparison to traditional dietary proxies such as foraging observation or
155 analysis of gut/scat contents (Dalerum and Angerbjörn 2005). Plasma proteins incorporate diet’s
156 isotopic signatures very rapidly, whereas bone collagen incorporates it very slowly and therefore

157 averages the composition of assimilated diets over a much longer time (Hobson and Clark 1992).
158 Thus, temporally segregated measurements of the same tissue in the same individual or
159 comparing differences between isotopic measurements on different tissues with contrasting
160 isotopic incorporation rates among individuals can reveal temporal changes in resource use
161 (Phillips and Eldridge 2006). Bolnick et al. (2003) and Bearhop et al. (2004) suggested that
162 variance in delta space among and within individuals may be useful proxies for niche breadth
163 and individual and population level specialization. Comparing the isotopic composition of fast
164 and slow tissues can also generate information about “grain size” of foraging animals (sensu
165 MacArthur and Levins 1964). Fine-grained foragers use resources in quick succession and hence
166 the isotopic composition of fast and slow tissues should be similar. In contrast, coarse-grained
167 foragers specialize temporally on a single resource and hence the isotopic composition of a fast
168 tissue should differ from that of a slow tissue, which integrates inputs over a long time scale.

169

170 **The limitations of the isotopic niche**

171 In a similar fashion to Hutchinson’s n-dimensional hyperspace with environmental
172 variables as coordinates, the isotopic niche is defined by a set of isotopic composition
173 measurements in a space with delta values as coordinates. The isotopic niche has many uses, but
174 it also has numerous limitations. Using it to make ecological inferences demands that we
175 recognize what we can and what we cannot infer from it.

176 Depicting isotopic measurements in delta space is intuitively appealing and informative
177 (Fig. 3). By plotting data of both resources and consumers in the same space, one can make
178 inferences about a) the potential contribution of each source to the consumers, b) the amount of
179 mixing of sources, and c) the contribution of variation among sources to variation in the

180 consumers' composition (Phillips and Gregg 2003 and references within), assuming that all the
181 relevant food sources have been characterized. Although one can learn much about an
182 organism's niche from the hypervolume that it occupies in delta space, isotopic niches have two
183 limitations: 1) they can be myopic, and 2) they can give deceptive estimates of niche width.
184 These limitations are worth recognizing.

185 Isotopic niches can be myopic for two reasons. The first one is that isotopic
186 measurements can only distinguish among resources with contrasting isotopic compositions and
187 blur the distinction among sources with similar compositions. Stable isotopes can tell us much
188 about the physiological pathways and status of resources (Dawson et al. 2002), but it is not
189 always possible to determine the specific taxonomic identity to sources. The myopic nature of
190 isotopic measurements can apply to both bionomic and scenopoetic axes. Wunder et al. (2005)
191 have emphasized the difficulties one faces when attempting to assign migrating birds to a precise
192 geographical breeding area. Stable isotopes are effective tools to study animal movements, but
193 they can have low accuracy (Rubenstein and Hobson 2004).

194 The second reason for the isotopic niche's myopic nature stems from the inconsistency of
195 isotopic incorporation. Macromolecules (i.e., carbohydrates, proteins, lipids) derived from diet,
196 and the elements from which they are constructed, undergo recombination and sorting during
197 digestion, metabolism, and tissue synthesis (reviewed by Martínez del Rio and Wolf 2005). The
198 inconsistency of isotopic incorporation, however, can be useful. The difference in $\delta^{15}\text{N}$ between
199 a consumer's tissues and its diet (denoted by $\Delta^{15}\text{N}$) has been very widely used to diagnose
200 trophic level (reviewed by Post 2002). The logic of this application is that if one knows the $\delta^{15}\text{N}$
201 of primary producers and one assumes that $\Delta^{15}\text{N}$ is constant across each trophic level, then, one
202 can estimate an animal's trophic level from its $\delta^{15}\text{N}$ composition, which is a fundamental variable

203 in defining an animal's niche (Post 2002). While there is little doubt that consumers' tissues are
204 enriched ^{15}N relative to resources, trophic enrichment can vary depending on physiology and
205 environmental factors (McCutchan et al. 2003). Until we have a better understanding of the
206 factors that determine the magnitude of $\Delta^{15}\text{N}$ (see Robbins et al. 2005, Martínez del Rio and
207 Wolf 2005), the use of the $\delta^{15}\text{N}$ axis of the isotopic niche will not provide an absolute measure of
208 trophic level, but is still useful in determining the relative trophic position of species within a
209 community.

210 Niche-theorists have proposed the dispersion in the distribution of points in niche space
211 as an estimate of niche width (Bolnick et al. 2002). It is natural (albeit misleading) to assume that
212 similar dispersion of points in delta space is evidence of a broad niche (Matthews and Mazumder
213 2004). For example, Bolnick et al. (2003) interpret "unexpectedly large isotopic differences
214 between individuals" as evidence of a high inter-individual component to niche width. This
215 interpretation is problematic because the processes that create variation in the isotopic
216 composition of producers can lead to widely divergent values. Dispersion in delta space is
217 dependent on the distance between the isotopic values of the alternative producers. Animals that
218 feed on two resources with widely divergent isotopic compositions will always be found to have
219 broader niches than animals that feed on food sources with less divergent delta values (Fig. 4),
220 but this may not always accurately reflect the true niche breadth. In the final section we will
221 describe how a metric of niche width that does not depend on the distance between the isotopic
222 values of producers can be constructed.

223

224 **Applications of the isotopic niche**

225 Many animals experience ontogenetic niche shifts (West et al. 2003). These shifts can be related
226 to changes in bionomic and/or scenopoetic factors and thus can be detected by SIA. Perhaps the
227 earliest use of SIA to study ontogenetic niche shifts was the application of $\delta^{15}\text{N}$ values to explore
228 the biochemical effects of nursing in humans and their offspring (Fogel et al. 1989). This
229 approach has now been used to assess the relative timing and nature of weaning in a growing list
230 of mammals (Newsome et al. 2006 and references there). Other vertebrate applications include
231 the use of SIA to examine the correlation between growth rate and diet composition in juveniles
232 (Snover 2002, Post 2003). SIA has also been utilized to assess ontogenetic changes in diet type
233 and/or quality in invertebrates, where in some cases, adult diets are nutritionally inadequate to
234 support observed juvenile growth (Hentschel 1998).

235 The identification of niche shifts, ontogenetic or otherwise, by SIA can also have
236 important conservation implications. For example, SIA demonstrated that loggerhead turtles
237 (*Caretta caretta*) use of productive, nearshore oceanic habitats not only increases juvenile
238 growth rates but may also increase by-catch risk (Snover 2002). Ecologists have also used
239 isotopes to document subtle niche shifts in lake trout (*Salvelinus namaycush*), which were
240 otherwise undetectable, following the invasion of two exotic bass species (Vander Zanden et al.
241 1999). SIA-derived scenopoetic and/or bionomic niche information can also be coupled with
242 toxicological data and satellite tracking technologies to identify the sources and vectors of
243 contaminants that threaten population viability (Finkelstein et al. 2006). Furthermore, SIA-
244 derived information on habitat preference(s) and connectivity within and among populations
245 could be combined with epidemiological data to identify disease vectors, especially for species

246 that have an inherently high potential for relatively fast transmission rates across spatial areas of
247 epidemic proportion (i.e., West Nile virus; Marra et al. 2004).

248 A third area of research where SIA-derived niche information continues to inform
249 conservation biology is in historic ecology, which aims to determine the true range of ecological
250 flexibility of species that may have experienced significant truncations in behavior due to direct
251 or indirect human disturbance (i.e., hunting, habitat loss). For example, SIA has been used to
252 identify differences in the use of coastal versus inland habitats by modern and ancient California
253 condor (*Gymnogyps californianus*) populations (Fig. 3B; Chamberlain et al. 2005, Fox-Dobbs et
254 al. 2006). These studies contend that conservation goals should emphasize the reintroduction of
255 condors (obligate scavengers) to coastal areas where populations would have access to stranded
256 marine mammal carcasses. Another study found a difference in the trophic level of historic
257 versus contemporary marbled murrelets (*Brachyramphus marmoratus*) in central California,
258 suggesting that recent decreases in large, energetically superior prey populations due to
259 overfishing is contributing to poor murrelet reproduction and recent population declines (Becker
260 and Beissinger 2006). The continual use of SIA to identify past versus present differences in
261 biogenic or scenopoetic niche space provides a means of describing the natural history of
262 species on ecologically and evolutionarily-relevant timescales, thus providing a means of
263 evaluating the significance of current ecological trends that is vital for the success of long-term
264 conservation and management strategies.

265

266 **Transforming from δ -space to p-space**

267 The degree of specialization and generalization in individuals and populations can inform
268 problems as diverse as the evolution of resource use (Bolnick 2003), the success of invading

269 exotics (Holt et al. 2005), and the processes that shape the composition of ecological
270 communities (Wiens and Graham 2005). Ecologists have devised a variety of metrics to assess
271 niche variation and the relative contribution of individual variation to these metrics (reviewed by
272 Bolnick et al. 2002). One can assess variation in the isotopic niche, but in a previous section we
273 identified one of the problems of isotopic niches as depicted in delta spaces. The variation within
274 and among individuals in isotopic composition is strongly dependent on how different the
275 isotopic signatures of the food sources are. An alternative to using δ -values *per se* to define
276 isotopic niches is to use mixing models to transform them into dietary proportions (p) of
277 different isotopic sources. Briefly, if one measures the isotopic composition of n elements, one
278 can determine the contribution of n+1 isotopically distinct sources by solving a system of n+1
279 linear equations (Fig. 5; see Phillips and Gregg 2001 for details). This transformation from δ -
280 space to p-space resolves the scaling discrepancies in δ -space discussed above, and permits using
281 the niche-width metrics most commonly used by ecologists (see Bolnick 2002). We hasten to
282 point out that depictions of the isotopic niche in δ -space and p-space are complementary rather
283 than alternative. By transforming data from delta-space to p-space, we gain the ability to
284 construct metrics of variation that are independent of the absolute value of isotopic signatures
285 and that are comparable to those of other niche formulations. However, we lose the insights on
286 the types of resources and locations in isoscapes that are revealed by δ -spaces.

287 Because mixing models are central tools in the analysis of isotopic niches, it is important
288 to pay attention to their assumptions and potential limitations. Both the isotopic composition of
289 isotopic sources and that of animal tissues are measured with variation. Consequently, the
290 numerical manipulations required to transform from δ -values to p-values involves error

291 propagation. Phillips and Gregg (2001) provide formulas for calculating variances, standard
292 errors (SE), and confidence intervals for p values. Using correct tissue-to-diet discrimination
293 factors is also important when estimating p values (Phillips and Gregg 2001). Finally, recall that
294 a mixing model resolves $n+1$ distinct sources if one measures n isotopes. Thus, a particular set of
295 δ -values may not define a point in p-space unless the number of distinct isotopic sources is
296 limited to one more than the number of δ -values measured. Phillips and Gregg (2003) have
297 devised a method that relaxes this requirement and makes it possible to determine the minimum
298 and maximum utilization of each source that is consistent with isotopic mass balance even when
299 one measures n isotopes and the number of resources exceeds $n+1$. However, the degree of
300 utilization within these bounds cannot be determined exactly but only as a range of possible
301 values (Phillips and Gregg 2003). In such cases, mixing models may only transform a δ -space
302 into a blurry p-space.

303

304 **Concluding remarks**

305 Scientific concepts sometimes lie dormant until new methodologies transform them and
306 revitalize them. Systems biology received intense interest from biologists in the 1960s and then
307 waned. Fertilized by the growth of the “omics” (genomics, proteonomics, metabolomics) and
308 fueled by the power of ever-faster computers, systems biology has reincarnated into a vigorous
309 field (Wolkenhauer 2001). In a similar fashion, the revival of the niche is the result of rapid
310 progress in bioinformatics and in the development of new technologies. Just as researchers
311 interested in systems biology and in tracking the evolution of biological systems rely on nucleic
312 acids and the polymerase chain reaction (PCR), ecologists interested in measuring the fluxes of
313 energy and materials among components of ecological systems increasingly rely on SIA (Yakir

314 2002). We predict the rapid growth of niche studies and contend that they will be stimulated by
315 faster, cheaper, and more accurate stable isotope analyses. Isotopic ecology will become an
316 important axis in the resurgent study of ecological niches.

317

318

319

320

321 **Acknowledgements**

322 Merav Ben David kindly gave us the data set used to draft figure 2a. CMR was funded by a
323 National Science Foundation grant (IBN-0110416). The information in this document has been
324 funded in part by the U.S. Environmental Protection Agency. It has been subjected to the
325 Agency's peer and administrative review, and approved for publication as an EPA document.
326 Mention of trade names or commercial products does not constitute endorsement or
327 recommendation for use. We thank Joe Shannon for a constructive review of the manuscript.

328

329

330

331

332

333

334

335

336

337

338 **References**

339 Bearhop S, Adams CE, Waldron S, Fuller RA, and Macleod H. 2004. Determining trophic niche
340 width: a novel approach using stable isotope analysis. *J Anim Ecol* 73: 1007-1012.

341 Becker BH and Beissinger SR. 2006. Centennial decline in the trophic level of an endangered
342 seabird after fisheries decline. *Conserv Biol* 20(2): 470-479.

343 Ben-David M, Flynn RW and Schell DM. 1997. Annual and seasonal changes in diets of
344 martens: evidence from stable isotopes. *Oecologia* 111: 280-291.

345 Bocher P, Cherel Y and Hobson KA. 2000. Complete trophic segregation between South
346 Georgian and common diving petrels during breeding at Iles Kerguelen. *Mar Ecol Prog
347 Ser* 208: 249-264.

348 Bolnick DI, Yang LH, Fordyce JA, Davis JM and Svanbäck R. 2002. Measuring individual-level
349 resource specialization. *Ecology* 83: 2936-2941.

350 Bolnick DI, Svanback R, Fordyce JA, Yang LH, Davis JM, Hulsey CD and Forister ML. 2003.
351 The ecology of individuals: Incidence and implications of individual specialization. *Am
352 Nat* 161: 1-28.

353 Bowen GJ and Revenaugh J. 2003. Interpolating the isotopic composition of modern meteoric
354 precipitation. *Water Resour Res* 39: 1299-1312.

355 Chamberlain CP, Waldbauer JR, Fox-Dobbs K, Newsome SD, Koch PL, Smith DR, Church ME,
356 Chamberlain SD, Sorenson KJ, and Risebrough R. 2005. Pleistocene to recent dietary
357 shifts in California condors. *Proc National Acad Sci* 102: 16707-16711.

358 Chase JM and Leibold MA. 2003. Ecological niches: linking classical and contemporary
359 approaches. Chicago, IL: University of Chicago Press.

360

361 Collins SL, Bettencourt LMA, Hagberg A, Brown RF, Moore MI, Bonito G, Delin KA, Jackson
362 SP, Johnson DW, Burleigh SC, Woodrow RR, and McAuley JM. 2006. New
363 opportunities in ecological sensing using wireless sensor networks. *Frontiers Ecol*
364 *Environ* 4: 402-406.

365 Dalerum F, and Angerbjörn A. 2005. Resolving temporal variation in vertebrate diets using
366 naturally occurring stable isotopes. *Oecologia* 144: 647-658.

367 Dawson TE, Mambelli S, Plamboeck AH, Templer PH, and Tu KP. 2002. Stable isotopes in
368 plant ecology. *Ann Rev Ecol Syst* 33: 507-559.

369 Elith J, Graham CH, Anderson RP, Dudík M, Ferrier S, Guisan A, Hijmans RJ, Huettmann F,
370 Leathwick JR, Lehmann A, Li J, Lohmann LG, Loiselle BA, Manion G, Moritz C,
371 Nakamura M, Nakazawa Y, Overton JM, Peterson AT, Phillips SJ, Richardson KS,
372 Scachetti-Pereira R, Schapire RE, Soberón, J, Williams, S, Wisz MS and Zimmermann
373 NE. 2006. Novel methods improve prediction of species' distributions from occurrence
374 data. *Ecography* 29: 129 -151.

375 Farquhar GD, Ehleringer JR and Kubick KT. 1989. Carbon isotope discrimination and
376 photosynthesis. *Ann Rev Plant Physiol Plant Mol Biol* 40: 503-537.

377 Finkelstein M, Keitt BS, Croll DA, Tershy BR, Jarman WM, Rodriguez S, Anderson DJ and
378 Sievert PR. 2006. Albatross species demonstrate regional differences in North Pacific
379 marine contamination. *Ecol Appl* 16(2): 678-686.

380 Fogel ML, Tuross N and Owsley DW. 1989. Nitrogen isotope tracers of human lactation in
381 modern and archaeological populations. Annual Report of the Director, Geophysical
382 Laboratory, Carnegie Institution of Washington.

383

384 Fox-Dobbs K, Stidham TA, Bowen GJ, Emslie SD, and Koch PL. 2006. Dietary controls on
385 extinction versus survival among avian megafauna in the late Pleistocene. *Geology* 34(8):
386 685-688.

387 Fry B. 2006. Stable isotope ecology. New York, NY: Springer.

388 Genner, MJ, Turner GF, Barker S and Hawkins SJ. 1999. Niche segregation among Lake Malawi
389 cichlid fishes? Evidence from stable isotope signatures. *Ecology Letters* 2; 185-190.

390 Grinnell J. 1917. The niche-relationships of the California thrasher. *Auk* 34: 427-433.

391 Hentschel BT. 1998. Intraspecific variations in $\delta^{13}\text{C}$ indicate ontogenetic diet changes in deposit-
392 feeding polychaetes. *Ecology* 79(4): 1357-1370.

393 Hobson KA, and Clark RG. 1992. Assessing avian diets using stable isotopes I: turnover of ^{13}C
394 in tissues. *Condor* 94:181-188.

395 Holt RD, Barfield M, and Gomulkiewicz R. 2005. Theories of niche conservatism and evolution:
396 could exotic species be potential tests? In: Sax D, Stachowicz J, and Gaines SD (Eds.)
397 Species Invasions: insights into ecology, evolution, and biogeography. Sunderland, MA:
398 Sinauer Associates.

399 Hubbell SP. 2001. The unified neutral theory of species abundance and diversity. Princeton, NJ:
400 Princeton University Press.

401 Hutchinson GE. 1957. Concluding remarks. Cold Spring Harbor Symp. *Quant Biol* 22: 415-427.

402 Hutchinson GE. 1978. An introduction to population biology. New Haven, CT: Yale University
403 Press.

404 Kohzu A, Kato C, Iwata T, Kishi D, Murakami M, Nakano N and Wada E. 2004. Stream food
405 web fueled by methane-derived carbon. *Aquat Microb Ecol* 36: 189-194.

406

407 Leibold MA. 1995. The niche concept revisited: mechanistic models and community context.

408 *Ecology* 76: 1371-1382.

409 MacArthur R and Levins R. 1964. Competition habitat selection and character displacement in a

410 patchy environment. *Proc Natl Acad Sci* 51: 1207-1210.

411 Marra PP, Griffing S, Cafree CL, Kilpatrick AM, McLean R, Brand C, Kramer L, and Novak R.

412 2004 West Nile virus and wildlife. *Bioscience* 54: 393-402.

413 Martinez del Rio C and Wolf BO. 2005. Mass-balance models for animal-isotopic ecology. In:

414 Stack M and Wang T (Eds.). Physiological and ecological adaptations to feeding in

415 vertebrates. Enfield, NH. Science Publishers.

416 Matthews B and Mazumder A. 2004. A critical evaluation of intrapopulation variation of delta

417 C-13 and isotopic evidence of individual specialization. *Oecologia* 140, 361-371.

418 McCutchan JH, Lewis WM, Jr., Kendall C, and McGrath CC. 2003. Variation in the trophic shift

419 for stable isotope ratios of carbon, nitrogen, and sulfur. *Oikos* 102: 378-390.

420 Newsome SD, Etnier MA, Aurioles-Gamboa D, and Koch PL. 2006. Using carbon and nitrogen

421 isotope values to investigate maternal strategies in northeast Pacific otariids. *Mar*

422 *Mammal Sci* 22(3): 556-572.

423 Phillips DL and Gregg JW. 2001. Uncertainty in source partitioning using stable isotopes.

424 *Oecologia* 127: 171-179.

425 Phillips DL and Gregg JW. 2003. Source partitioning using stable isotopes: coping with too

426 many sources. *Oecologia* 136: 261-269.

427 Phillips DL and Eldridge PM. 2006. Estimating the timing of diet shifts using stable isotopes.

428 *Oecologia* 147: 195-203.

429 Post DM. 2002. Using stable isotopes to estimate trophic position: models, methods, and
430 assumptions. *Ecology* 83: 703-718.

431 Post DM. 2003. Individual variation in the timing of ontogenetic niche shifts in largemouth bass.
432 *Ecology* 84(5): 1298-1310.

433 Radtke RL, Lenz P, Showers W, and Moksness E. Environmental information stored in otoliths:
434 insights from stable isotopes. *Mar Biol* 127: 161-170.

435 Robbins CT, Felicetti LA, and Sponheimer M. 2005. The effect of dietary protein quality on
436 nitrogen isotope discrimination in mammals and birds. *Oecologia* 144:534-540

437 Robinson D. 2001. $\delta^{15}\text{N}$ as an integrator of the nitrogen cycle. *Trends Ecol Evol* 16: 153-162.

438 Rubenstein DR and Hobson KA. 2004. From birds to butterflies: animal movement patterns and
439 stable isotopes. *Trends Ecol Evol* 19: 256-263.

440 Snover ML. 2002. Estimation of age, detection of habitat shifts, and the implications of growth
441 rate variability on population dynamics for loggerhead and Kemp's ridley sea turtles.
442 (PhD dissertation) Durham, NC: Duke University.

443 Tilman D. 1988. Plant strategies and the dynamics and structure of plant communities. Princeton,
444 NJ: Princeton University Press.

445 Vander Zanden MJ, Casselman JM, and Rasmussen JB. 1999. Stable isotope evidence for the
446 food web consequence of species invasions in lakes. *Nature* 401(6752): 464-467.

447 Wassenaar LI and Hobson KA. 2000. Stable carbon and hydrogen isotope ratios reveal breeding
448 origins of red-winged blackbirds. *Ecol Appl* 10(3): 911-916.

449 West JB, Bowen GJ, Cerling TE, and Ehleringer JR. 2006. Stable isotopes as one of nature's
450 recorders. *Trends Ecol Evol* 21: 408-414.

451 West MJ, King PL and White DJ. The case for developmental ecology. *Anim Behav* 66: 617-622.

452 Wiens JJ and Graham CH. 2005. Niche conservatism: integrating evolution, ecology, and
453 conservation biology. *Ann Rev Ecol Evol Syst* 36: 519-539.

454 Wolf BO and Martinez del Rio C. 2004. How important are CAM succulents as sources of water
455 and nutrients for desert consumers? A review. *Isotop in Environ Health Sci* 39: 53-67.

456 Wolkenhauer O. 2001. Systems biology: the reincarnation of systems theory applied to biology.
457 *Briefings in Bioinformatics* 2: 258-270.

458 Wunder MB, Kester CL, Knopf FL and Rye R. 2005. A test of geographic assignment using
459 isotope tracers in feathers of known origin. *Oecologia* 144: 607-617.

460 Yakir D. 2002. Global enzymes: sphere of influence. *Nature* 416: 795.

461

462

463

464

465

466

467

468

469

470

471

472

473

474

475

TABLE 1

Gradient	Isotope System	High δ -Values	Low δ -Values	Scenopoetic	Bionomic
Trophic Level	$\delta^{13}\text{C} / \delta^{15}\text{N}$	High Levels	Low Levels		√
C ₃ – C ₄ Vegetation	$\delta^{13}\text{C}$	C ₄ Plants	C ₃ Plants		√
Marine – Terrestrial	$\delta^{15}\text{N} / \delta^{13}\text{C} / \delta^{34}\text{S}$	Marine	Terrestrial	√	√
Latitude (Terrestrial)	$\delta^2\text{H} / \delta^{18}\text{O}$	Low Latitudes	High Latitudes	√	
Latitude (Marine)	$\delta^{13}\text{C} / \delta^{15}\text{N}$	Low Latitudes	High Latitudes	√	
Altitude	$\delta^{13}\text{C}$	High Altitudes	Low Altitudes	√	
Altitude	$\delta^2\text{H}$	Low Altitudes	High Altitudes	√	
Inshore – Offshore	$\delta^{13}\text{C}$	Inshore	Offshore	√	
Benthic – Pelagic	$\delta^{13}\text{C}$	Benthic	Pelagic	√	√
Aridity	$\delta^{13}\text{C} / \delta^{15}\text{N}$	Xeric	Mesic/Hydric	√	
Eutrophication	$\delta^{15}\text{N}$	Polluted	Pristine	√	
Temperature	$\delta^{18}\text{O}$	Cooler	Warmer	√	
Geologic Substrate	$\delta^{87}\text{Sr}$	Young Rocks	Old Rocks	√	
Oxic – Anoxic	$\delta^{15}\text{N} / \delta^{13}\text{C} / \delta^{34}\text{S}$	Oxic	Anoxic	√	
Methanogenesis	$\delta^{13}\text{C}$	Photosynthetic	Methanogenic	√	

476

477

478

479

480

481

482

483

484

485

486

487

488

489

490 **FIGURE 1.** Isotopic ratios are typically expressed as the ratio of the heavy (H) to light (L)
491 isotope and converted into delta notation (δ -values) through comparison of sample isotope ratios
492 to ratios of internationally accepted standards. Standards for common systems include Vienna-
493 Pee Dee Belemnite limestone (V-PDB) for carbon, atmospheric N₂ for nitrogen, and VSMOW
494 for hydrogen and oxygen. The units are expressed as parts per thousand or per mil (‰).

495

496 **FIGURE 2.** Geographical patterns in the δ D and δ^{18} O of precipitation have been used widely to
497 track animal movements and study population connectivity, thus supplying information on
498 scenopoetic factors of the ecological niche.

499

500 **FIGURE 3.** Two examples of how delta-space can supply information on the bionomic and
501 scenopoetic axes of the ecological niche. In some cases, an isotopic axis can have both bionomic
502 and scenopetic components (panel 2), where feeding on a marine or terrestrial food web implies
503 inhabiting a marine/terrestrial habitat. Data from Wassenaar and Hobson (2000) and
504 Chamberlain et al. (2005).

505

506 **FIGURE 4.** Variance in delta-space is dependent on the isotopic composition of resources. The
507 variance in δ^{13} C in the larvae of the marsh beetle (*Helodidae*, panel b) is 29 times greater than
508 that of American marten (*Martes americana*, panel b. When δ^{13} C values are transformed to p
509 values and the variances are recalculated, the values for these two species are roughly similar.
510 Data from Kohzu et al. (2004) and Ben-David et al. (1997).

511

512

513 **FIGURE 5.** Transforming from d- to p-space requires solving a system of 3 linear equations in 3
514 unknowns for each point. The figure illustrates the transformation from delta- to p-space for 3
515 species that rely on intertidal, freshwater, and/or terrestrial food-webs. The points in p space are
516 represented in a ternary diagram.

517

518

519

520

521

522

523

524

525

526

527

528

529

530

531

532

533

534

535

536

538

FIGURE 1

540

Isotope Ratio	Delta (δ) Notation Derivation	δ -Value
$^H X/^L X$	$\left[\left(\frac{^H X/^L X_{sample}}{^H X/^L X_{standard}} - 1 \right) \times 1000 \right]$	$\delta^H X$
$^{13}C/^{12}C$	$\left[\left(\frac{^{13}C/^{12}C_{sample}}{^{13}C/^{12}C_{standard}} - 1 \right) \times 1000 \right]$	$\delta^{13}C$

542

544

546

547

548

549

550

551

552

553

554

555

556

557

558

559

560

561

562

563

564

565

566

567

568

569

570

571

572

573

574

575

576

577

578

579

580

581

582

584

FIGURE 2

586

588

590

592

594

596

598

600

602

604

606

608

610

612

614

616

618

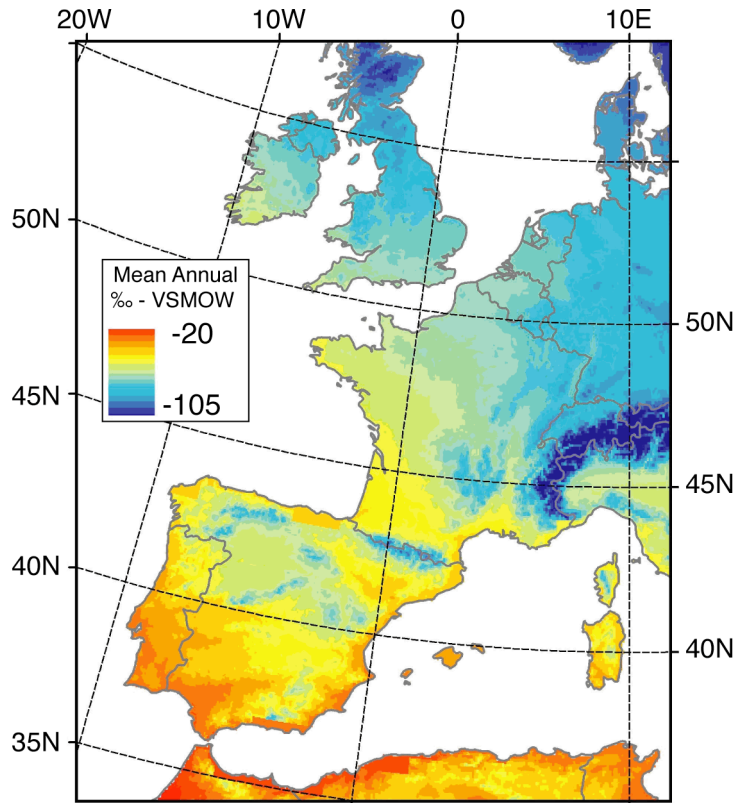
619

620

621

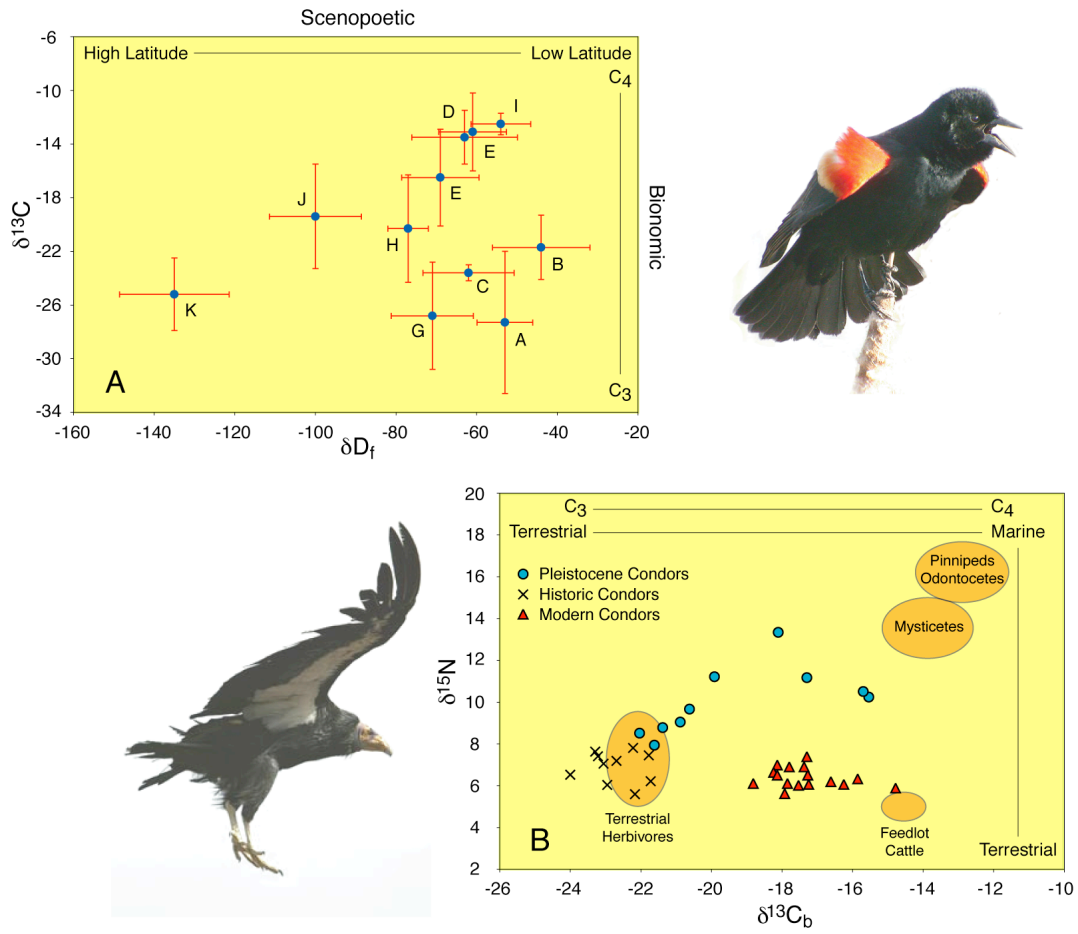
622

623



625
627
629
631
633
635
637
639
641
643
645
647
649
651
652
653
654
655
656
657
658
659
660

FIGURE 3



662

FIGURE 4

664

666

668

670

672

674

676

678

680

682

684

686

688

690

692

694

696

697

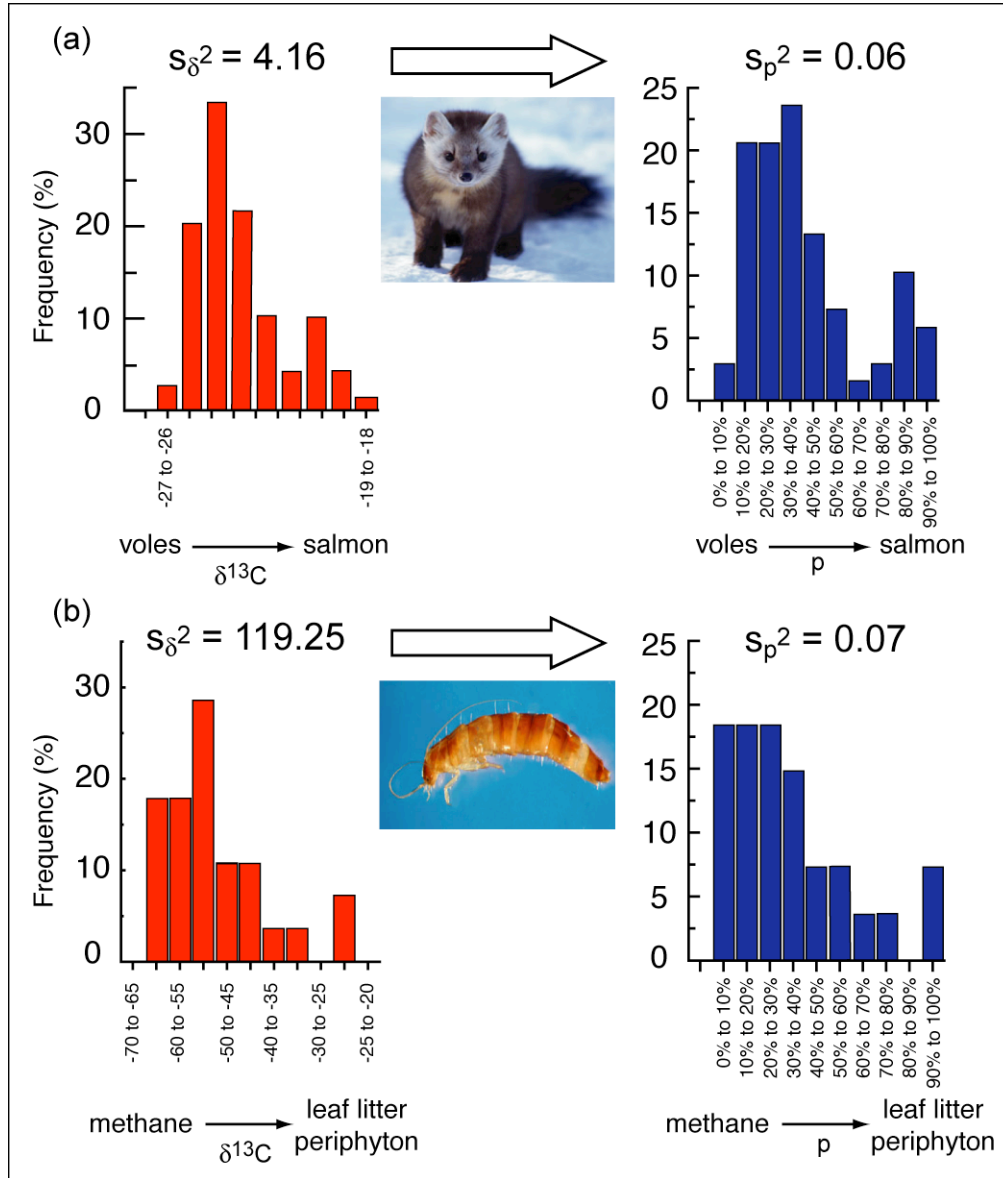
698

699

700

701

702



704

FIGURE 5

706

708

